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## ORIGINAL ARTICLE

# Relationships between retinal layer thickness and brain volumes in the UK Biobank cohort

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## Abstract

**Background and purpose:** Current methods to diagnose neurodegenerative diseases are costly and invasive. Retinal neuroanatomy may be a biomarker for more neurodegenerative processes and can be quantified in vivo using optical coherence tomography (OCT), which is inexpensive and noninvasive. We examined the association of neuroretinal morphology with brain MRI image-derived phenotypes (IDPs) in a large cohort of healthy older people.

**Methods:** UK Biobank participants aged 40 to 69 years old underwent comprehensive examinations including ophthalmic and brain imaging assessments. Macular retinal nerve fibre layer (mRNFL), macular ganglion cell-inner plexiform layer (mGCIPL), macular ganglion cell complex (mGCC) and total macular thicknesses were obtained from OCT. Magnetic resonance imaging (MRI) IDPs assessed included total brain, grey matter, white matter and hippocampal volume. Multivariable linear regression models were used to evaluate associations between retinal layers thickness and brain MRI IDPs, adjusting for demographic factors and vascular risk factors.

**Abbreviations:** AD, Alzheimer disease; BMI, body mass index; CI, confidence interval; IDP, image-derived phenotype; IOP, intraocular pressure; IOPcc, corneal-compensated intraocular pressure; logMAR, logarithm of the minimum angle of resolution; MCI, mild cognitive impairment; mGCC, macular ganglion cell complex; mGCIPL, macular ganglion cell-inner plexiform layer; mGCL, macular ganglion cell layer; mIPL, macular inner plexiform layer; MRI, magnetic resonance imaging; mRNFL, macular retinal nerve fibre layer; pRNFL, peripapillary retinal nerve fibre layer; SD-OCT, spectral-domain optical coherence tomography; TABS, Topcon Advanced Boundary Segmentation.

Anthony P. Khawaja and Praveen J. Patel are senior authors taking joint credit and responsibility.

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**Results:** A total of 2131 participants (mean age 55 years; 51% women) with both gradable OCT images and brain imaging assessments were included. In multivariable regression analysis, thinner mGCIPL, mGCC and total macular thickness were all significantly associated with smaller total brain ( $p < 0.001$ ), grey matter and white matter volume ( $p < 0.01$ ), and grey matter volume in the occipital pole ( $p < 0.05$ ). Thinner mGCC and total macular thicknesses were associated with smaller hippocampal volume ( $p < 0.02$ ). No association was found between mRNFL and the MRI IDPs.

**Conclusions:** Markers of retinal neurodegeneration are associated with smaller brain volumes. Our findings suggest that retinal structure may be a biomarker providing information about important brain structure in healthy older adults.

#### KEYWORDS

brain MRI markers, cognitive impairment, optical coherence tomography, retinal layers, retinal neurodegeneration

## INTRODUCTION

Neurodegenerative diseases are emerging as the foremost challenge for biomedical science in the 21st century. It has been estimated that 46 million people are living with dementia, and this number is expected to rise to 131 million by 2050 [1,2]. Prevalence of dementia increases with age, affecting 11%, 32% and 82% of people aged over 65, 75 and 85 years, respectively [3]. Some projections suggest that, due to population aging, the prevalence of Alzheimer disease (AD), the most common form of dementia, may triple by 2050 [2,3]. Studies have reported age-related decreases in global and regional brain changes measured by magnetic resonance imaging (MRI) [4]. Brain volume declines with age at a rate of around 5% per decade after age 40, and the rate of decline accelerates after 70 years of age [5,6]. Brain MRI is accurate and sensitive in detecting structural variations in the brain, some of which are associated with neurodegeneration, but is relatively costly and time consuming [7]. Simpler, resource saving, reproducible biomarkers would offer the prospect of enabling earlier diagnosis of dementia.

The retina and optic nerve share their embryological origin with the brain, and are widely regarded as part of the central nervous system [8]. Retinal microvasculature and neuronal components offer a unique window on tissues that are closely allied to intracranial structures [9]. Consequently, the eye is vulnerable to similar processes that are associated with neurodegenerative diseases [8]. Spectral-domain optical coherence tomography (SD-OCT) is an in vivo imaging tool that allows noninvasive, high-resolution examination of retinal structure [10]. Automated segmentation techniques now make quantitative assessment of retinal sublayers a viable proposition [11]. This offers the prospect of retinal imaging contributing to the diagnosis and monitoring of diseases characterised by structural changes in the brain.

There is a well-recognised link between dementia and degenerative changes in the retina and optic nerve [12]. Both histopathological and clinical studies have shown that evidence of retinal ganglion cell degeneration in AD patients [13–16], but only the Rotterdam

Study has evaluated whether retinal degeneration may be a marker for subclinical brain disease [17]. To date, most interest has focused on the thickness of the peripapillary retinal nerve fibre layer (pRNFL) as a potential biomarker for dementia [18]. Few studies have examined the relationship between retinal sublayer thickness and cerebral atrophy on MRI [17,19]. In this UK Biobank study, we aimed to investigate the association between inner retinal layer thicknesses and regional brain volumes in a healthy population to better understand how accessible and affordable eye measurement may contribute as a biomarker for brain structure.

## METHODS

### Study population

UK Biobank is a very large community-based cohort of UK residents registered with the National Health Service and aged 40 to 69 years at enrolment. Baseline examinations were carried out between 2006 and 2010 at 22 study assessment centres. The North West Multicentre Research Ethics Committee approved the study in accordance with the principles of the Declaration of Helsinki. The overall study protocol (<http://www.ukbiobank.ac.uk/resources/>) and protocols for individual tests (<http://biobank.ctsu.ox.ac.uk/crystal/docs.cgi>) are available online. In brief, participants answered a wide-ranging touch-screen questionnaire covering demographic, socioeconomic and lifestyle information. Physical measures included blood pressure, height and weight. Body mass index (BMI) was defined as weight divided by height squared.

### Ocular measurements

In late 2009, eye measurements were conducted in six assessment centres by trained staff following standard operating procedures; detailed methods have been published [20,21]. Visual acuity and

intraocular pressure (IOP) were measured. Corneal-compensated IOP ( $IOP_{cc}$ ) was measured with the ocular response analyser (Reichert, Philadelphia, PA, USA) to examine the influence that corneal biomechanical characteristics might have on IOP measures [20,22].

### Spectral-domain optical coherence tomography

Spectral-domain optical coherence tomography imaging was performed using the Topcon 3D OCT-1000 Mk2 (Topcon, Tokyo, Japan). Image acquisition was performed under mesopic conditions, without pupillary dilation using the three-dimensional macular volume scan (512 horizontal A scans per B scan; 128 B scans in a  $6 \times 6$  mm<sup>2</sup> raster pattern). The Topcon Advanced Boundary Segmentation (TABS) algorithm was used for automated segmentation [23]. Retinal layers were labelled as macular retinal nerve fibre layer (mRNFL), macular ganglion cell-inner plexiform layer (mGCIPL), macular ganglion cell complex (mGCC) and total macular thickness. Thickness of the mGCIPL includes the macular ganglion cell layer (mGCL) and macular inner plexiform layer (mIPL), whereas mGCC includes both mRNFL and mGCIPL. We followed the Advised Protocol for OCT Study Terminology and Elements (APOSTEL) guidelines, except with respect to the total macular thickness, which we define as the thickness from the inner limiting membrane to the retinal pigment epithelium [24]. mRNFL is composed of axons of the retinal ganglion cell, whereas mGCL is composed of ganglion cell bodies, and mIPL is composed of retinal ganglion cell dendrites. We used the average thickness of the retinal layers in the macula across nine retinal sub-fields in a 6-mm-diameter circle centred at the true fovea location, as derived from the ETDRS (Early Treatment Diabetic Retinopathy Study) [25]. We have added the prefix "m" to the sublayer abbreviations to denote as macular measures, as some SD-OCT metrics can be derived from images centred on the optic disc.

### MRI Brain Image-Derived Phenotypes (IDPs)

MRI imaging protocols were designed by the UK Biobank Imaging Working Group (<http://www.ukbiobank.ac.uk/expert-working-groups>). Details of the image acquisition and processing are available on the UK Biobank website in the brain scan protocol (<http://biobank.ctsu.ox.ac.uk/crystal/refer.cgi?id=2367>) and brain imaging documentation ([http://biobank.ctsu.ox.ac.uk/crystal/docs/brain\\_mri.pdf](http://biobank.ctsu.ox.ac.uk/crystal/docs/brain_mri.pdf)). Briefly, brain imaging was carried out using a single standard Siemens Skyra 3T scanner with a 32-channel radio-frequency (RF) receive head coil. Key acquisition parameters for each modality are summarised in a prior publication [26]. The T1-weighted scans were acquired using three-dimensional magnetization-prepared rapid gradient-echo (MPRAGE) (resolution 1 mm<sup>3</sup> isotropic voxels) and analysed with the Functional Magnetic Resonance Imaging of the Brain (FMRIB) Software Library (FSL) (<http://fsl.fmrib.ox.ac.uk/fsl>). Further details on MRI acquisition and analysis have been reported

elsewhere [26,27]. Brain images were analysed to derive multiple distinct individual measures of brain structure and function, known as brain image-derived phenotypes (IDPs) [26]. Total brain volume was defined as the sum of grey matter volume and white matter volume. Grey matter volume in the frontal, temporal and occipital poles and hippocampal volume were averaged from the right and left volumes. Total brain, grey matter and white matter volumes were normalised for head size. Grey-matter volume in the frontal, temporal and occipital poles (V1/V2) and hippocampus volume were normalised for head size by multiplying the individual brain IDPs by the volumetric scaling from T1 head image to standard space. Brain IDPs were normalised for head size, which is a close proxy for intracranial volume [28]. Normalised brain IDPs were termed as total brain, grey matter, white matter and hippocampal normalised.

### Inclusion and exclusion criteria

To ensure the accuracy of retinal thickness assessment, we have excluded OCT scans of poor quality consistent with the OSCAR IB" [(O)= obvious problems including violation of the protocol; (S) poor signal strength defined as  $\geq 15$  dB; (C) wrong centration of scan; (A) algorithm failure; (R) retinal pathology other than MS related; (I) illumination; and (B) beam placement]. (OSCAR-IB) criteria [29]. In general, the OSCAR-IB criteria requires good quality OCT scans and signal strength with no visible retinal pathology. Among those with SD-OCT data, we excluded participants who withdrew their consent, had poor SD-OCT signal strength, had an image quality score  $<45$ , had poor centration certainty or had poor segmentation certainty using TABS software [30,31]. Participants with the following conditions were also excluded from the study: visual acuity worse than 0.5 logarithm of the minimum angle of resolution (logMAR) (approximately 6/18 on Snellen chart),  $IOP_{cc}$  of  $<6$  mm Hg or  $>24$  mm Hg [32], self-reported ocular disorders or diseases, self-reported diabetes or self-reported neurodegenerative disease. This group of participants were excluded from the study because of the well-recognised impact these conditions have on retinal layer thickness [33–35].

### Statistical analysis

The present analysis was based on cross-sectional data. For this analysis, if both eyes of a patient were eligible for inclusion, one eye was randomly selected using Stata software (version 16; StataCorp, College Station, TX, USA). The z scores of the four retinal sublayer thickness and brain MRI markers were calculated by subtracting the mean value from the value of the observation and dividing by the standard deviation. We examined the association of retinal sublayer thickness (explanatory or independent variable) with brain MRI markers (dependent variable) using two multivariable linear regression models: Model 1 adjusted for age, age squared and sex; and Model 2 additionally adjusted for race, education, BMI, mean arterial blood pressure and smoking status. Cardiovascular risk factors

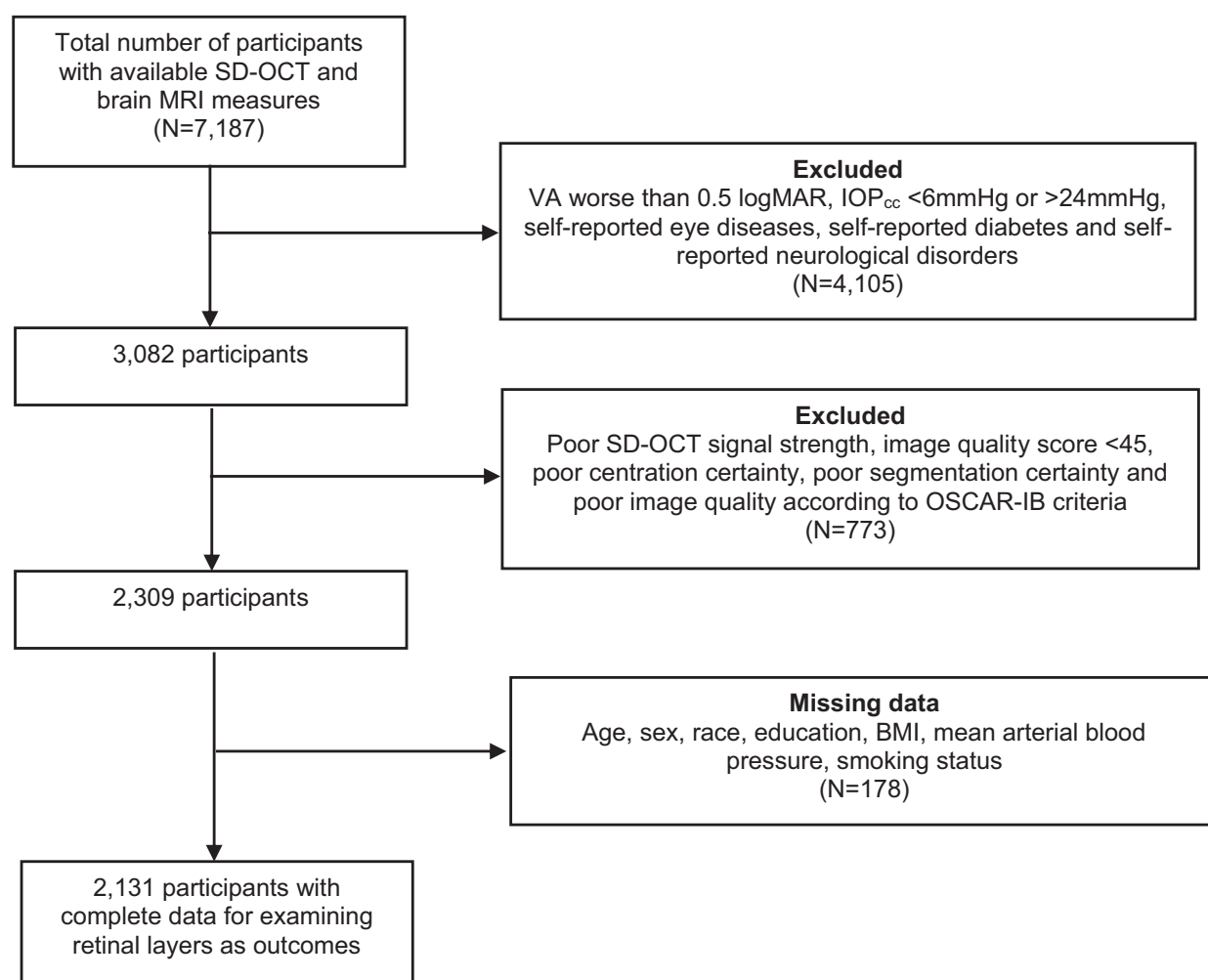
including BMI, mean arterial blood pressure and smoking status were adjusted for in the multivariable models in view of their relationship with brain MRI markers [36–38] and retinal layers [39]. We minimised the effect of age on cerebral atrophy by adjusting for age and age squared. In sensitivity analysis, we additionally adjusted for total grey matter normalised when the outcome was hippocampal normalised. The  $\beta$  coefficients represent standardised mean difference in z scores of total brain, grey or white matter and hippocampal normalised per SD decrease in retinal sublayer thickness. We applied linear transformation and have used z scores as standardizing scores to facilitate the interpretation of the value of the individual brain IDPs.

## RESULTS

There were 7187 participants with data on SD-OCT macular imaging and brain MRI scans. Of these, 4105 people with visual acuity worse than 0.5 logMAR (equivalent to 6/18 on Snellen chart), IOP<sub>cc</sub>

of <6 mm Hg or >24 mm Hg, self-reported eye diseases, diabetes and neurological disorders were excluded. Of the remaining 3082, 773 participants with poor SD-OCT image quality were excluded, and 178 participants were further excluded due to missing data for covariables. There were complete data (age, sex, race, education, BMI, mean arterial blood pressure and smoking) for 2131 of these participants (Figure 1).

The characteristics of the study population are shown in Table 1. The mean age was 54.6 years (SD = 7.4), and 1084 (51%) of the participants were female. The overall mean (SD) of the different retinal layers was: mRNFL (29.2  $\mu$ m [SD, 4.6  $\mu$ m]), mGCIPL (74.1  $\mu$ m [5.9  $\mu$ m]), mGCC (103.3  $\mu$ m [8.0  $\mu$ m]) and total macular thickness (278.7  $\mu$ m [12.7  $\mu$ m]). Table 2 shows the multivariable analysis of retinal sublayer thickness with total brain, grey matter, white matter and hippocampal normalised after adjusting for age, age squared and sex. In general, thinner mRNFL, mGCIPL, mGCC and total macular thickness were all significantly associated with smaller total brain, grey matter and white matter normalised (except the association between mRNFL and total white matter normalised). Reduced mRNFL,



**FIGURE 1** Flowchart of participants included in the study. BMI, body mass index; IOP<sub>cc</sub>, corneal-compensated intraocular pressure; logMAR, logarithm of the minimum angle of resolution; MRI, magnetic resonance imaging; SD-OCT, spectral-domain optical coherence tomography; VA, visual acuity.

**TABLE 1** Characteristics of the study population (*n* = 2131)

Characteristic	N	Mean $\pm$ SD <sup>a</sup>	%
Age, years	2131	54.6 $\pm$ 7.4	
Sex, %			
Male	1047		49.1
Female	1084		50.9
Race, %			
White	2063		96.8
Non-White	68		3.2
Education level, %			
O level and less	600		28.2
Professional qualification or A level	553		25.9
Degree and above	978		45.9
Body mass index, kg/m <sup>2</sup>	2131	26.7 $\pm$ 4.2	
Smoking status, %			
Never	1370		64.3
Former	640		30.0
Current	121		5.7
Mean arterial blood pressure, mm Hg	2131	100.7 $\pm$ 12.1	

<sup>a</sup>Mean (SD) for continuous variables and percentages for categorical variables.

mGCC and total macular thickness were associated with smaller hippocampal normalised, but the associations were not significant ( $p > 0.05$ ) when we additionally adjusted for total grey matter normalised.

Table 3 shows the multivariable analysis of retinal sublayer thickness with MRI IDPs after adjusting for age, age squared, sex, race, education, BMI, mean arterial blood pressure and smoking status. Thinner mGCIPL, mGCC, and total macular thickness were all significantly associated with smaller total brain normalised, standardised mean difference (95% CI) per SD decrease in retinal sublayer thickness, respectively:  $-0.08$  (95% confidence interval [CI]:  $-0.12, -0.05$ ;  $p < 0.001$ ),  $-0.08$  (95% CI:  $-0.12, -0.04$ ;  $p < 0.001$ ) and  $-0.08$  (95% CI:  $-0.11, -0.04$ ;  $p < 0.001$ ). Reduced mGCIPL, mGCC and total macular thickness were significantly associated with grey matter normalised, standardised mean difference (95% CI) per SD decrease in retinal sublayer thickness, respectively:  $-0.04$  (95% CI:  $-0.08, -0.01$ ;  $p = 0.009$ ),  $-0.05$  (95% CI:  $-0.08, -0.02$ ;  $p = 0.003$ ) and  $-0.06$  (95% CI:  $-0.09, -0.02$ ;  $p = 0.001$ ). Thinner mGCIPL, mGCC and total macular thickness were also significantly associated with white matter normalised, standardised mean difference (95% CI) per SD decrease in retinal sublayer thickness, respectively:  $-0.09$  (95% CI:  $-0.13, -0.05$ ;  $p < 0.001$ ),  $-0.08$  (95% CI:  $-0.12, -0.04$ ;  $p < 0.001$ ) and  $-0.07$  (95% CI:  $-0.11, -0.03$ ;  $p = 0.001$ ). Reduced mGCC thickness was associated with smaller grey matter normalised in the frontal pole, standardised mean difference (95% CI) per SD decrease in mGCC thickness:  $-0.04$  (95% CI:  $-0.08, -0.002$ ;  $p = 0.038$ ). Thinner

mGCIPL, mGCC and total macular thickness were associated with smaller grey matter normalised in the occipital pole, standardised mean difference (95% CI) per SD decrease in retinal sublayer thickness, respectively:  $-0.06$  (95% CI:  $-0.10, -0.02$ ;  $p = 0.003$ ),  $-0.06$  (95% CI:  $-0.10, -0.02$ ;  $p = 0.006$ ) and  $-0.05$  (95% CI:  $-0.09, -0.008$ ;  $p = 0.020$ ). Thinner mGCC and total macular thickness were associated with smaller hippocampal normalised, standardised mean difference (95% CI) per SD decrease in retinal sublayer thickness, respectively:  $-0.05$  (95% CI:  $-0.09, -0.008$ ;  $p = 0.018$ ), and  $-0.05$  (95% CI:  $-0.09, -0.01$ ;  $p = 0.010$ ) but were no longer significant ( $p > 0.05$ ) when we additionally adjusted for total grey matter normalised. There were no significant associations between mRNFL thickness and the brain MRI measures. Additional adjustment for IOP<sub>cc</sub>, the sole modifiable risk factor for glaucoma, did not meaningfully change the effect estimates between retinal sublayer thicknesses and brain MRI markers.

## DISCUSSION

In this study, we identified that thinner mGCIPL, mGCC and total macular thickness were associated with smaller total brain normalised, grey and white matter normalised, and grey matter normalised in the occipital pole. Reduced mGCC was associated with smaller grey matter normalised in the frontal pole, whereas mRNFL thickness was not associated with the MRI IDPs. This suggests that retinal thickness measures provide not only useful information on brain function, but also on anatomically relevant brain structures.

Cognitive impairment in dementia is characterised by cerebral atrophy. Our study has shown that thinner mGCIPL, mGCC and total macular thicknesses were associated with smaller total brain, grey and white matter normalised. In addition, because of the age-related effects on cerebral atrophy, we adjusted for age and age squared to minimise the possibility of confounding by age. Our data suggest that neuronal damage may occur in the retina and throughout the brain. Our result was consistent with a report from the Rotterdam Study Group that reported thinner inner retinal layers were associated with total brain, grey and white matter volumes [17]. Our findings showed that thinner mGCIPL, mGCC and total macular thicknesses were associated with reduced grey-matter normalised in the occipital pole. It is well known that atrophy of the occipital lobe, particularly V1/V2, is known to cause inner retinal layer atrophy by retrograde transsynaptic axonal degeneration. In agreement with our findings, Ong et al. reported that thinner mGCIPL was associated with smaller grey matter volume in the occipital lobe [19]. These data are also consistent with a number of studies in control subjects and patients with multiple sclerosis [40–42]. The association of inner retinal layer atrophy was strongest for V1 and V2 using validated segmentation software. Taken together, the published data suggest a strong association between pathology and atrophy in these areas. The question becomes more complex for impaired higher visual function. AD pathology of the lateral occipital cortices and parietal lobes are not known to be associated with atrophy of

**TABLE 2** Age- and sex-adjusted estimated change in brain MRI markers per standard deviation decrease in average mRNFL, mGCIPL, mGCC and total macular thickness

	mRNFL thickness			mGCIPL thickness			mGCC thickness			Total macular thickness		
	$\beta$	95% CI	p value	$\beta$	95% CI	p value	$\beta$	95% CI	p value	$\beta$	95% CI	p value
Total brain normalised	-0.04	(-0.07, -0.002)	<b>0.04</b>	-0.08	(-0.12, -0.05)	<b>&lt;0.001</b>	-0.08	(-0.12, -0.05)	<b>&lt;0.001</b>	-0.08	(-0.12, -0.05)	<b>&lt;0.001</b>
Grey matter normalised												
Total normalised	-0.04	(-0.07, -0.005)	<b>0.022</b>	-0.05	(-0.08, -0.01)	<b>0.007</b>	-0.06	(-0.09, -0.02)	<b>0.001</b>	-0.06	(-0.09, -0.03)	<b>&lt;0.001</b>
Frontal pole	-0.02	(-0.06, 0.01)	0.19	-0.04	(-0.07, 0.0008)	0.06	-0.04	(-0.08, -0.004)	<b>0.030</b>	-0.04	(-0.08, -0.003)	<b>0.034</b>
Temporal pole	0.002	(-0.04, 0.04)	0.92	-0.03	(-0.07, 0.01)	0.21	-0.02	(-0.06, 0.02)	0.39	-0.04	(-0.08, 0.006)	0.09
Occipital pole	-0.02	(-0.06, 0.02)	0.42	-0.06	(-0.10, -0.02)	<b>0.004</b>	-0.06	(-0.10, -0.01)	<b>0.009</b>	-0.05	(-0.09, -0.009)	<b>0.017</b>
White matter volume												
Total normalised	-0.02	(-0.06, 0.02)	0.28	-0.09	(-0.13, -0.05)	<b>&lt;0.001</b>	-0.08	(-0.12, -0.04)	<b>&lt;0.001</b>	-0.07	(-0.11, -0.03)	<b>0.001</b>
Hippocampal normalised	-0.04	(-0.08, -0.004)	<b>0.030</b>	-0.03	(-0.07, 0.005)	0.09	-0.05	(-0.09, -0.01)	<b>0.012</b>	-0.05	(-0.09, -0.01)	<b>0.008</b>

Note: Values represent standardised mean difference in z scores of total brain, grey or white matter, and hippocampal normalised per SD decrease in retinal sublayer thickness. Values are adjusted for age, age squared and sex. Bold values denote statistical significance at the  $p < 0.05$  level.

Abbreviations: mGCC, macular ganglion cell complex; mGCIPL, macular ganglion cell-inner plexiform layer; mRNFL, macular retinal nerve fibre layer.

**TABLE 3** Multivariable adjusted estimated change in brain MRI markers per standard deviation decrease in average mRNFL, mGCIPL, mGCC and total macular thickness

	mRNFL thickness			mGCIPL thickness			mGCC thickness			Total macular thickness		
	$\beta$	95% CI	p value	$\beta$	95% CI	p value	$\beta$	95% CI	p value	$\beta$	95% CI	p value
Total brain normalised	-0.03	(-0.07, 0.004)	0.08	-0.08	(-0.12, -0.05)	<b>&lt;0.001</b>	-0.08	(-0.12, -0.04)	<b>&lt;0.001</b>	-0.08	(-0.11, -0.04)	<b>&lt;0.001</b>
Grey matter normalised												
Total normalised	-0.03	(-0.06, 0.002)	0.06	-0.04	(-0.08, -0.01)	<b>0.009</b>	-0.05	(-0.08, -0.02)	<b>0.003</b>	-0.06	(-0.09, -0.02)	<b>0.001</b>
Frontal pole	-0.02	(-0.06, 0.01)	0.24	-0.04	(-0.07, 0.001)	0.06	-0.04	(-0.08, -0.002)	<b>0.038</b>	-0.04	(-0.08, 0.0001)	0.05
Temporal pole	0.005	(-0.04, 0.05)	0.80	-0.02	(-0.07, 0.02)	0.23	-0.02	(-0.06, 0.03)	0.46	-0.03	(-0.07, 0.007)	0.11
Occipital pole	-0.02	(-0.06, 0.02)	0.37	-0.06	(-0.10, -0.02)	<b>0.003</b>	-0.06	(-0.10, -0.02)	<b>0.006</b>	-0.05	(-0.09, -0.008)	<b>0.020</b>
White matter normalised												
Total normalised	-0.02	(-0.06, 0.02)	0.32	-0.09	(-0.13, -0.05)	<b>&lt;0.001</b>	-0.08	(-0.12, -0.04)	<b>&lt;0.001</b>	-0.07	(-0.11, -0.03)	<b>0.001</b>
Hippocampal normalised	-0.04	(-0.08, 0.0002)	0.05	-0.03	(-0.07, 0.006)	0.09	-0.05	(-0.09, -0.008)	<b>0.018</b>	-0.05	(-0.09, -0.01)	<b>0.010</b>

Note: Values represent standardised mean difference in z scores of total brain, grey or white matter and hippocampal normalised per SD decrease in retinal sublayer thickness. Values are adjusted for age, age squared, sex, race, education, body mass index, mean arterial blood pressure and smoking. Bold values denote statistical significance at the  $p < 0.05$  level.

Abbreviations: mGCC, macular ganglion cell complex; mGCIPL, macular ganglion cell-inner plexiform layer; mRNFL, macular retinal nerve fibre layer.



inner retinal layers in the macula. Therefore, a limitation of the study is that impaired visual perception in AD is unlikely to be captured by atrophy of the mGCC/mGCIPL.

Hippocampal atrophy is a major feature of AD and a diagnostic marker for AD at the mild cognitive impairment stage [43]. Histological studies have shown considerable neuronal loss in the hippocampus at onset of symptoms [44]. In agreement with histological findings, longitudinal studies reported accelerated hippocampal volume loss in AD [45–48] and mild cognitive impairment (MCI) [49,50]. In addition to smaller hippocampal volumes observed in patients with cognitive impairment, evidence also suggests that the inhibition of adult hippocampal neurogenesis leads to memory impairments [51,52], whereas enhancing neurogenesis improves memory performance [53,54]. Given that hippocampal atrophy is a prominent feature of AD and MCI, our findings that, in an adult population, thinner mGCC and total macular thicknesses were associated with smaller hippocampal normalised, suggests that common mechanisms may lead to hippocampal and retinal atrophy. Additionally, we examined the association between total macular thickness and brain MRI, and observed that thinner total macular thickness was associated with smaller brain MRI structural measures. Because the presence of imaging artefacts may decrease the accuracy of specific retinal thickness measurements, measurement of total macular thickness is more accurate compared to specific sublayer retinal thickness [55]. In line with our findings, the Rotterdam Study reported that thinner inner retinal layers were associated with smaller hippocampal volumes, although grey volume was not adjusted in the multivariable models [17].

Nonetheless, in a sensitivity analysis, we did not find an association between the inner retinal layers or total macular thickness and hippocampal normalised after the additional adjustment of total grey matter volume. This suggests that, at least among the healthy UK Biobank participants, the relationship between thinner retinal measures and smaller hippocampal volumes is explicable as a consequence of smaller total grey matter volume [56], which could be a consequence of either global neurodegeneration or differences in neurodevelopment. Measurement of relative rates of volume loss over time in the same individuals will disambiguate this question.

Previous studies by Blanks et al. [13] and Hinton et al. [14] reported that patients with AD have apparent histological signs of retinal ganglion cell and mRNFL loss compared to controls, consistent with accelerated neurodegeneration. Clinic-based studies have demonstrated that patients with AD or mild cognitive impairment have reduced pRNFL [34], mGCIPL [57], mGCC [58] and total macular thickness [59,60] compared to controls. AD is associated with cerebral atrophy on high-resolution MRI [43]. Although our findings suggest that inner retinal and total macular thickness measurements may be a biomarker for brain structure, further work would be needed to examine the utility of retinal parameters as a diagnostic or screening tool for diseases that affect brain structure.

We did not identify significant associations between mRNFL thickness and brain MRI structural measures. However, pRNFL was not measured in our study. Thinner pRNFL was associated with grey

matter volume in the temporal lobe in a Singapore study [19] and thinner pRNFL was associated with smaller grey matter and white matter volumes in the Rotterdam Study [17]. As the mRNFL axon bundles that arise from retinal ganglion cells converge toward the optic disc, mRNFL in the macula is much thinner compared to pRNFL (and vice versa for mGCIPL) [61], which may explain the lack of association between mRNFL thickness and brain MRI measures in our study. In addition, reduction in dendritic complexity and area occurs prior to retinal ganglion cell death and loss [62], which suggests that mGCIPL may be more sensitive at detecting neurodegenerative damage compared to mRNFL.

The strengths of our study include its large sample, the quantitative assessment of retinal sublayers on SD-OCT and the brain structures on MRI. Limitations of the study include the UK Biobank is a volunteer cohort, and participants are likely to be healthier than the general population. In addition, a large number of the participants with eye data did not yet have data on brain structural MRI measures. The self-reported nature of glaucoma, ocular disorders, diabetes or neurodegenerative disease also could result in misclassification bias. However, as both the retinal structures measured with SD-OCT and brain MRI structures are objective measures, the latter is more likely to be nondifferential misclassification and bias the effect estimates toward the null. The cross-sectional design of our study limits the ability to determine the cause and effect of the relationship between retinal structures and brain MRI markers. Among healthy older adults (i.e., our cohort), there is extreme variability in the relationship between hippocampal size and memory [63]. Our study may not be able to identify pathological trends, but rather provides information on the trends to be expected in a healthy, older population.

In conclusion, thinner mGCIPL, mGCC and total macular thickness are associated with smaller total brain, grey matter and white matter normalised in a cohort of healthy adults. Our findings support the concept that retinal layer thickness is a biomarker for processes affecting central nervous system structure. Further research examining the utility of retinal measures as an affordable, noninvasive investigation of neurological conditions is warranted.

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## CONFLICT OF INTEREST

Charles Reisman reports employment by Topcon Healthcare Solutions, Inc. outside the submitted work. Paul J. Foster reports personal fees from Allergan, Carl Zeiss, Google/DeepMind and Santen, a grant from Alcon, outside the submitted work. Praveen J. Patel reports grants from Topcon Inc., outside the submitted work. Anthony P. Khawaja reports personal fees from Aerie, Allergan, Google Health, Novartis, Thea and Santen, all outside the submitted work. Sharon Y. L. Chua, Axel Petzold, Nicholas G. Strouthidis, Gerassimos Lascaratos, Denize Atan, Bing Zhang, Stephen M. Smith, Paul M. Matthews and Peng T. Khaw declare no competing interests.

## AUTHORS' CONTRIBUTIONS

Peng T. Khaw and Paul J. Foster contributed to the conception and design of the study. Sharon Y. L. Chua and Paul J. Foster contributed to the data analyses, data interpretation and wrote the draft of the manuscript. All authors reviewed the results, and read and critically revised the manuscript. All authors approved the final manuscript.

## ETHICAL APPROVAL

The North West Multicenter Research Ethics Committee approved the study (reference no. 06/MRE08/65), in accordance with the tenets of the Declaration of Helsinki. Detailed information about the study is available at the UK Biobank website ([www.ukbiobank.ac.uk](http://www.ukbiobank.ac.uk))

## DATA AVAILABILITY STATEMENT

Researchers wishing to access UK Biobank data can register and apply at <https://www.ukbiobank.ac.uk/>.

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## REFERENCES

- GBD 2015 Neurological Disorders Collaborator Group. Global, regional, and national burden of neurological disorders during 1990–2015: a systematic analysis for the Global Burden of Disease Study 2015. *Lancet Neurol.* 2017;16(11):877–897.
- Prince M, Wimo A, Guerchet M, Ali G, Wu Y, Prina M. *World Alzheimer Report 2015: The Global Impact of Dementia - An analysis of prevalence, incidence, cost and trends.* Alzheimer's Disease International. London, UK: Alzheimer's Disease International; 2015.
- Hebert LE, Weuve J, Scherr PA, Evans DA. Alzheimer disease in the United States (2010–2050) estimated using the 2010 census. *Neurology.* 2013;80(19):1778–1783.
- Peters R. Ageing and the brain. *Postgrad Med J.* 2006;82(964):84–88.
- Scahill RI, Frost C, Jenkins R, Whitwell JL, Rossor MN, Fox NC. A longitudinal study of brain volume changes in normal aging using serial registered magnetic resonance imaging. *Arch Neurol.* 2003;60(7):989–994.
- Svennerholm L, Boström K, Jungbjer B. Changes in weight and compositions of major membrane components of human brain during the span of adult human life of Swedes. *Acta Neuropathol.* 1997;94(4):345–352.
- Ledig C, Schuh A, Guerrero R, Heckemann RA, Rueckert D. Structural brain imaging in Alzheimer's disease and mild cognitive impairment: biomarker analysis and shared morphometry database. *Sci Rep.* 2018;8(1):11258.
- Frost S, Martins R, Kanagasingam Y. Ocular biomarkers for early detection of Alzheimer's disease. *J Alzheimers Dis.* 2010;22(1):1–16.
- Liew G, Mitchell P, Wong T, et al. Retinal microvascular signs and cognitive impairment. *J Am Geriatr Soc.* 2009;57(10):1892–1896.
- Huang D, Swanson EA, Lin CP, et al. Optical coherence tomography. *Science (New York, NY).* 1991;254(5035):1178–1181.
- Ishikawa H, Stein D, Wollstein G, Beaton S, Fujimoto J, Schuman J. Macular segmentation with optical coherence tomography. *Invest Ophthalmol Vis Sci.* 2005;46(6):2012–2017.
- Hinton D, Sadun A, Blanks J, Miller C. Optic-nerve degeneration in Alzheimer's disease. *N Engl J Med.* 1986;315(8):485–487.
- Blanks JC, Hinton DR, Sadun AA, Miller CA. Retinal ganglion cell degeneration in Alzheimer's disease. *Brain Res.* 1989;501(2):364–372.
- Hinton DR, Sadun AA, Blanks JC, Miller CA. Optic-nerve degeneration in Alzheimer's disease. *N Engl J Med.* 1986;315(8):485–487.
- Blanks JC, Torigoe Y, Hinton DR, Blanks RHI. Retinal pathology in Alzheimer's disease. I. Ganglion cell loss in foveal/parafoveal retina. *Neurobiol Aging.* 1996;17(3):377–384.
- Iseri PK, Altinas O, Tokay T, Yuksel N. Relationship between cognitive impairment and retinal morphological and visual functional abnormalities in Alzheimer disease. *J Neuro-ophthalmol.* 2006;26(1):18–24.
- Mutlu U, Bonnemaier PWM, Ikram MA, et al. Retinal neurodegeneration and brain MRI markers: the Rotterdam study. *Neurobiol Aging.* 2017;60:183–191.
- Tsai C, Ritch R, Schwartz B, et al. Optic nerve head and nerve fiber layer in Alzheimer's disease. *Arch Ophthalmol.* 1991;109(2):199–204.
- Ong YT, Hilal S, Cheung CY, et al. Retinal neurodegeneration on optical coherence tomography and cerebral atrophy. *Neurosci Lett.* 2015;584:12–16.
- Chan MP, Grossi CM, Khawaja AP, et al. Associations with intraocular pressure in a large cohort: results from the UK Biobank. *Ophthalmology.* 2016;123(4):771–782.
- Cumberland PM, Bao Y, Hysi PG, et al. Frequency and distribution of refractive error in adult life: methodology and findings of the UK Biobank study. *PLoS ONE.* 2015;10(10):e0139780.
- Luce D. Methodology for cornea compensated IOP and corneal resistance factor for the Reichert ocular response analyzer. *Invest Ophthalmol Vis Sci.* 2006;47(13):2266–.
- Yang Q, Reisman CA, Wang Z, et al. Automated layer segmentation of macular OCT images using dual-scale gradient information. *Opt Express.* 2010;18(20):21293–21307.
- Cruz-Herranz A, Balk LJ, Oberwahrenbrock T, et al. The APOSTEL recommendations for reporting quantitative optical coherence tomography studies. *Neurology.* 2016;86(24):2303–2309.
- Early Treatment Diabetic Retinopathy Study Research Group. Grading diabetic retinopathy from stereoscopic color fundus photographs—an extension of the modified Airlie House classification. ETDRS report number 10. Early Treatment Diabetic Retinopathy Study Research Group. *Ophthalmology.* 1991;98(5 Suppl):786–806.
- Miller KL, Alfaro-Almagro F, Bangerter NK, et al. Multimodal population brain imaging in the UK Biobank prospective epidemiological study. *Nat Neurosci.* 2016;19(11):1523–1536.
- Alfaro-Almagro F, Jenkinson M, Bangerter NK, et al. Image processing and Quality Control for the first 10,000 brain imaging datasets from UK Biobank. *NeuroImage.* 2018;166:400–424.
- Smith SM, Zhang Y, Jenkinson M, et al. Accurate, robust, and automated longitudinal and cross-sectional brain change analysis. *NeuroImage.* 2002;17(1):479–489.

29. Tewarie P, Balk L, Costello F, et al. The OSCAR-IB consensus criteria for retinal OCT quality assessment. *PLoS ONE*. 2012;7(4):e34823.
30. Ko F, Foster PJ, Strouthidis NG, et al. Associations with retinal pigment epithelium thickness measures in a large cohort: results from the UK Biobank. *Ophthalmology*. 2017;124(1):105-117.
31. Patel PJ, Foster PJ, Grossi CM, et al. Spectral-domain optical coherence tomography imaging in 67 321 adults: associations with macular thickness in the UK Biobank study. *Ophthalmology*. 2016;123(4):829-840.
32. National Institute for Health and Care Excellence. Glaucoma: diagnosis and management. NICE guideline [NG81] 2017. <http://www.nice.org.uk/guidance/ng81>. (accessed 12/06/2020)
33. Vujosevic S, Midena E. Retinal layers changes in human pre-clinical and early clinical diabetic retinopathy support early retinal neuronal and Muller cells alterations. *J Diabetes Res*. 2013;2013:905058.
34. Thomson KL, Yeo JM, Waddell B, Cameron JR, Pal S. A systematic review and meta-analysis of retinal nerve fiber layer change in dementia, using optical coherence tomography. *Alzheimer's Dement (Amst)*. 2015;1(2):136-143.
35. Diniz-Filho A, Abe RY, Zangwill LM, et al. Association between intraocular pressure and rates of retinal nerve fiber layer loss measured by optical coherence tomography. *Ophthalmology*. 2016;123(10):2058-2065.
36. Friedman JI, Tang CY, de Haas HJ, et al. Brain imaging changes associated with risk factors for cardiovascular and cerebrovascular disease in asymptomatic patients. *JACC Cardiovasc Imaging*. 2014;7(10):1039-1053.
37. Launer LJ, Lewis CE, Schreiner PJ, et al. Vascular factors and multiple measures of early brain health: CARDIA brain MRI study. *PLoS ONE*. 2015;10(3):e0122138.
38. Roos AD, van der Grond J, Grong Jvd, Mitchell G, Westenberg J. Magnetic resonance imaging of cardiovascular function and the brain. *Circulation*. 2017;135(22):2178-2195.
39. Xu L, Zhou JQ, Wang S, et al. Localized retinal nerve fiber layer defects and arterial hypertension. *Am J Hypertens*. 2013;26(4):511-517.
40. Petzold A, Nijland PG, Balk LJ, et al. Visual pathway neurodegeneration winged by mitochondrial dysfunction. *Ann Clin Transl Neurol*. 2015;2(2):140-150.
41. Balk LJ, Steenwijk MD, Tewarie P, et al. Bidirectional trans-synaptic axonal degeneration in the visual pathway in multiple sclerosis. *J Neurol Neurosurg Psychiatry*. 2015;86(4):419-424.
42. Balk LJ, Twisk JW, Steenwijk MD, et al. A dam for retrograde axonal degeneration in multiple sclerosis? *J Neurol Neurosurg Psychiatry*. 2014;85(7):782-789.
43. Frisoni GB, Fox NC, Jack CR Jr, Scheltens P, Thompson PM. The clinical use of structural MRI in Alzheimer disease. *Nat Rev Neurol*. 2010;6(2):67-77.
44. Braak H, Braak E. Evolution of neuronal changes in the course of Alzheimer's disease. *J Neural Transm Suppl*. 1998;53:127-140.
45. Jack CR Jr, Petersen RC, Grundman M, et al. Longitudinal MRI findings from the vitamin E and donepezil treatment study for MCI. *Neurobiol Aging*. 2008;29(9):1285-1295.
46. Jack CR Jr, Shiung MM, Gunter JL, et al. Comparison of different MRI brain atrophy rate measures with clinical disease progression in AD. *Neurology*. 2004;62(4):591-600.
47. Fox NC, Black RS, Gilman S, et al. Effects of Abeta immunization (AN1792) on MRI measures of cerebral volume in Alzheimer disease. *Neurology*. 2005;64(9):1563-1572.
48. van de Pol LA, Hensel A, van der Flier WM, et al. Hippocampal atrophy on MRI in frontotemporal lobar degeneration and Alzheimer's disease. *J Neurol Neurosurg Psychiatry*. 2006;77(4):439-442.
49. Jack CR Jr, Shiung MM, Weigand SD, et al. Brain atrophy rates predict subsequent clinical conversion in normal elderly and amnesic MCI. *Neurology*. 2005;65(8):1227-1231.
50. van de Pol LA, van der Flier WM, Korf ES, Fox NC, Barkhof F, Scheltens P. Baseline predictors of rates of hippocampal atrophy in mild cognitive impairment. *Neurology*. 2007;69(15):1491-1497.
51. Shors TJ, Miesegaes G, Beylin A, Zhao M, Rydel T, Gould E. Neurogenesis in the adult is involved in the formation of trace memories. *Nature*. 2001;410(6826):372-376.
52. Goodman T, Trouche S, Massou I, et al. Young hippocampal neurons are critical for recent and remote spatial memory in adult mice. *Neuroscience*. 2010;171(3):769-778.
53. Kempermann G, Kuhn HG, Gage FH. Experience-induced neurogenesis in the senescent dentate gyrus. *J Neurosci*. 1998;18(9):3206-3212.
54. van Praag H, Kempermann G, Gage FH. Running increases cell proliferation and neurogenesis in the adult mouse dentate gyrus. *Nat Neurosci*. 1999;2(3):266-270.
55. Asrani S, Essaid L, Alder BD, Santiago-Turla C. Artifacts in spectral-domain optical coherence tomography measurements in glaucoma. *JAMA Ophthalmol*. 2014;132(4):396-402.
56. Nobis L, Manohar SG, Smith SM, et al. Hippocampal volume across age: Nomograms derived from over 19,700 people in UK Biobank. *Neuroimage Clin*. 2019;23:101904.
57. Cheung CY, Ong YT, Hilal S, et al. Retinal ganglion cell analysis using high-definition optical coherence tomography in patients with mild cognitive impairment and Alzheimer's disease. *J Alzheimer's Dis*. 2015;45(1):45-56.
58. Bayhan HA, Aslan Bayhan S, Celikbilek A, Tanik N, Gurdal C. Evaluation of the chorioretinal thickness changes in Alzheimer's disease using spectral-domain optical coherence tomography. *Clin Exp Ophthalmol*. 2015;43(2):145-151.
59. Garcia-Martin ES, Rojas B, Ramirez AI, et al. Macular thickness as a potential biomarker of mild Alzheimer's disease. *Ophthalmology*. 2014;121(5):1149.e3-1151.e3.
60. den Haan J, Verbraak FD, Visser PJ, Bouwman FH. Retinal thickness in Alzheimer's disease: a systematic review and meta-analysis. *Alzheimer's Dement (Amst)*. 2017;6:162-170.
61. Leung CK, Chan WM, Yung WH, et al. Comparison of macular and peripapillary measurements for the detection of glaucoma: an optical coherence tomography study. *Ophthalmology*. 2005;112(3):391-400.
62. Williams PA, Thirgood RA, Oliphant H, et al. Retinal ganglion cell dendritic degeneration in a mouse model of Alzheimer's disease. *Neurobiol Aging*. 2013;34(7):1799-1806.
63. Van Petten C. Relationship between hippocampal volume and memory ability in healthy individuals across the lifespan: review and meta-analysis. *Neuropsychologia*. 2004;42(10):1394-1413.

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